



ROSANA



R
O
S
A
N
A

Representation **O**f **S**timuli **A**s **N**eural **A**ctivity

UCM: Universidad Complutense de Madrid
UAM: Universidad Autonoma de Madrid
FhG-IBMT: Fraunhofer Institute for Biomedical Engineering
IBMC: Institute for Molecular and Cell Biology of Porto
UEx: Univ. of Exeter

“Perception”

“Internal Representation” of the external world

R
O
S
A
N
A

Understand the fundamentals of perception



Determine the principles of coding of sensory stimuli



Interactions sensory inputs - activity of CNS neurons



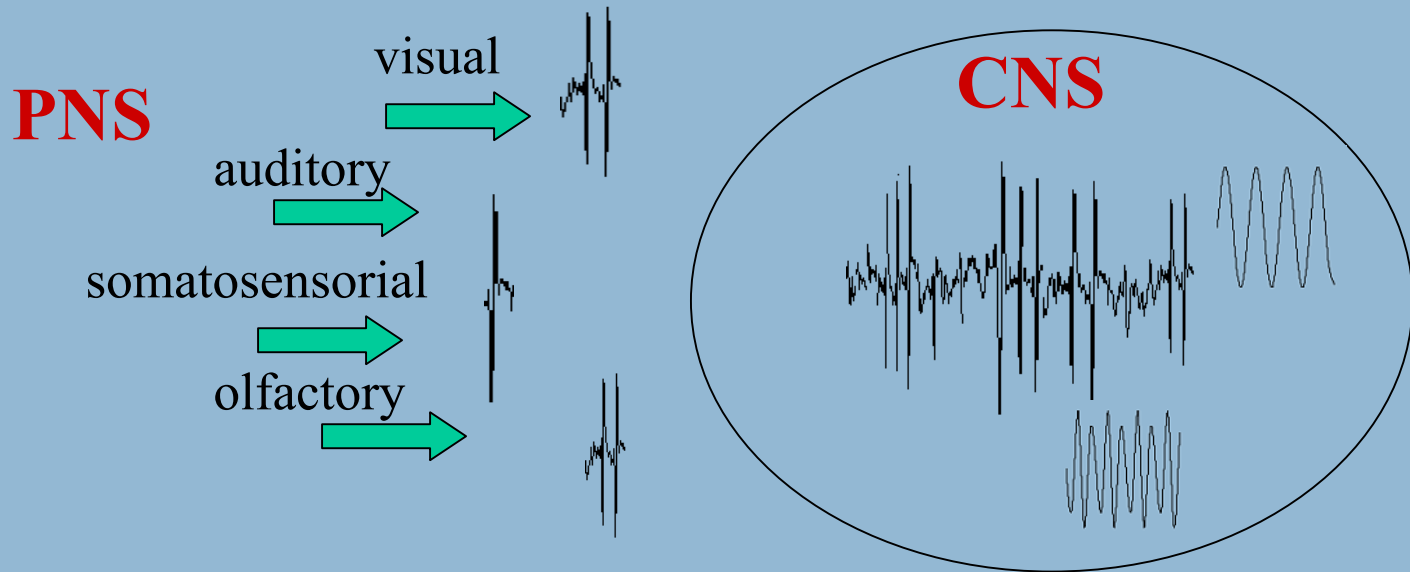
we will be able to reproduce the same spatiotemporal pattern of activity



“Perception”

Oscillations and Information processing

R
O
S
A
N
A



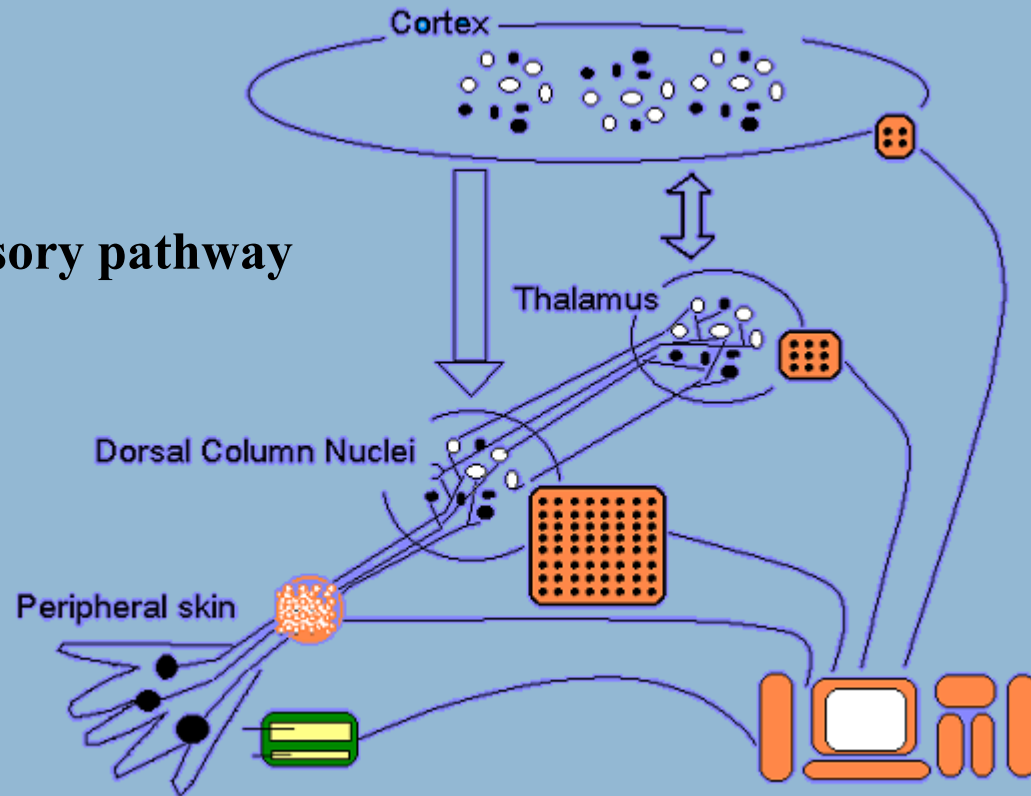
- Record the neural activity in PNS and CNS
- Develop mathematical models using experimental data



Our Experiment

R
O
S
A
N
A

Lemniscal sensory pathway



Methodology

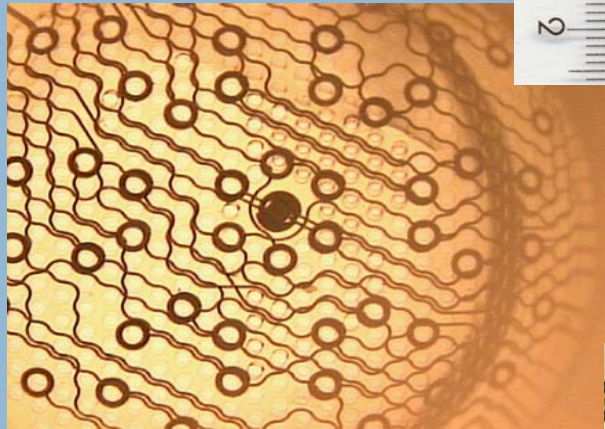
**R
O
S
A
N
A**

- ◆ Sieve microelectrodes
- ◆ Skin stimulator
- ◆ Sieve electrodes implantation
- ◆ Histological – quantification of nerve regeneration
- ◆ Simultaneous recordings from the PNS – CNS
- ◆ Data analysis
- ◆ Mathematical models

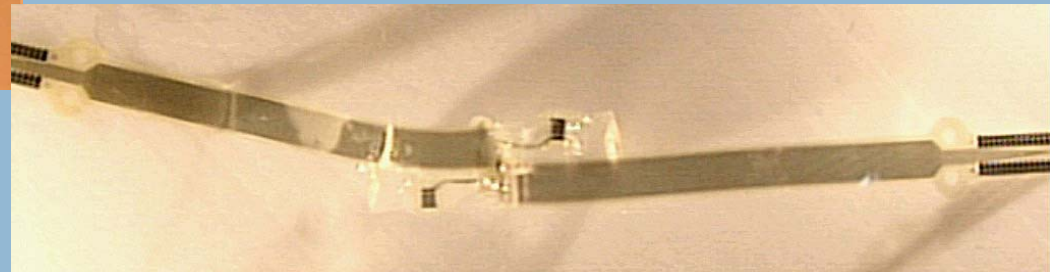
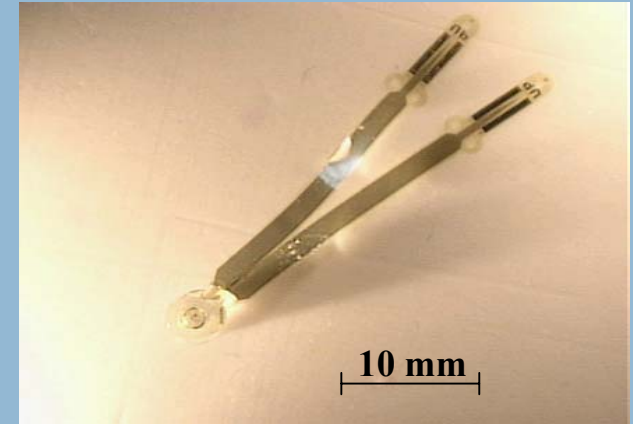


Sieve electrode design

R
O
S
A
N
A



500 μm

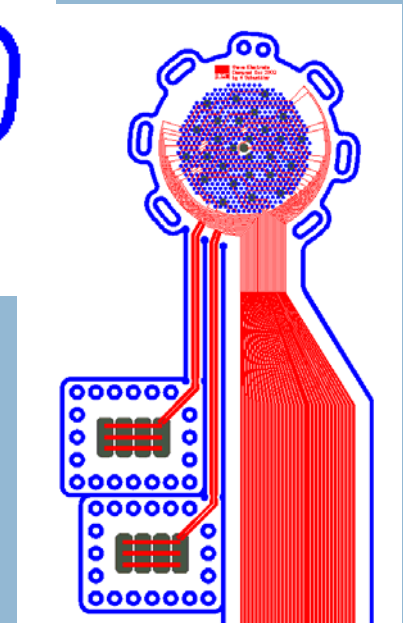
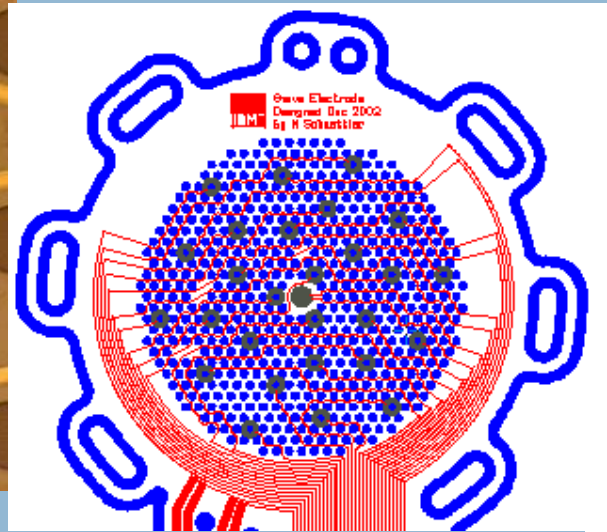
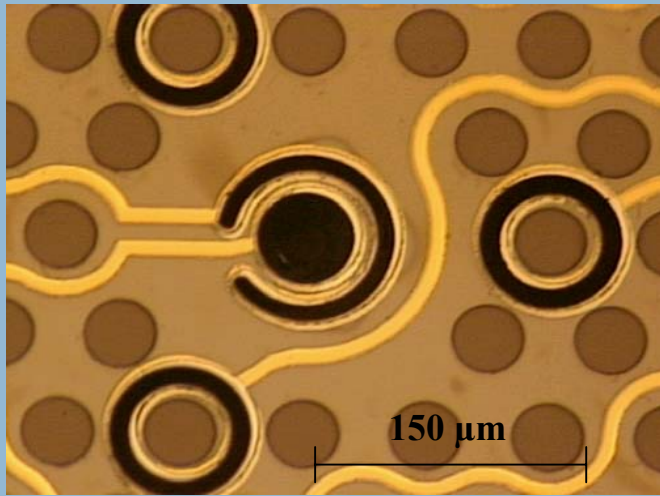


- 54 ring electrodes (50/90 μm)
- Electrodes material with platinum-black
- Easy to contact the electrodes
- Big stimulation counter electrodes distal and proximal
- 2 mm tubes as nerve channels



Sieve electrode design

R
O
S
A
N
A



- Improved mechanical fixation between electrode and silicone tube
- 570 regeneration holes (40 μm)
- Recording reference electrodes with no holes around the electrodes to reduce crosstalk
- Two possible sizes for recording reference (ring and circle)



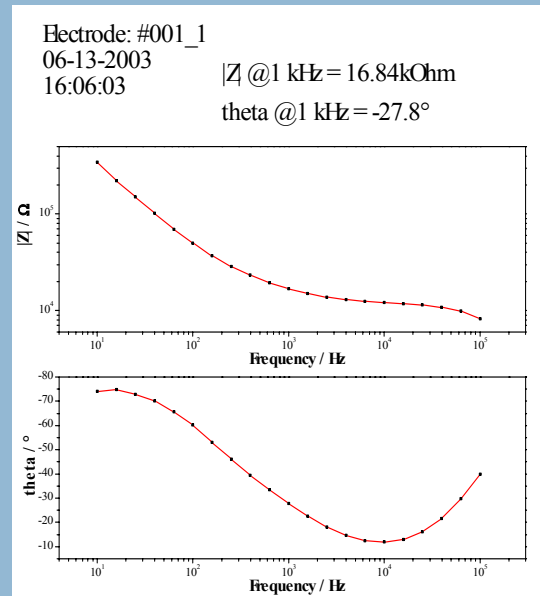
Electrode impedance for production and implantation controlling

R
O
S
A
N
A

Electrode impedance at 1 kHz (N=20)

Defect	Absolute Value / kOhm		Phase / °	
	Mean	SD	Mean	SD
Demolished	1196	26	-77	0,78
Line crack	2218	50	-89,3	1,0
No connector contact	44 510	26 443	-92	27
Wet connector	1,65	1,02	-23	9,4
Non defect	16,8	1,4	-28,7	2,9

Electrode Impedance



Electrode check:

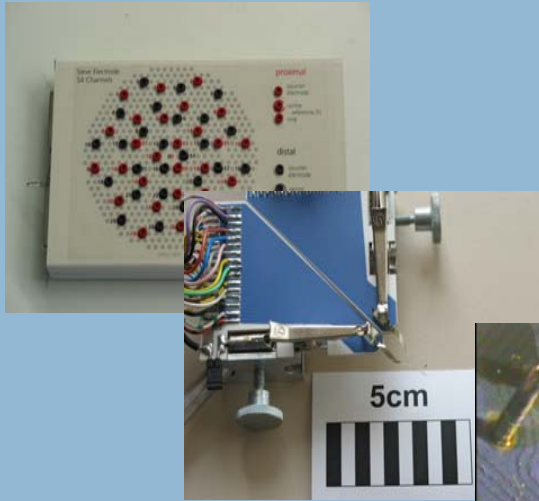


Electrode impedance have to be between
5 kOhm and 50 kOhm at 1 kHz

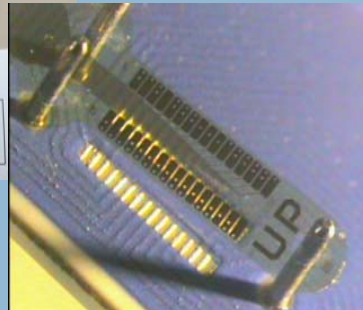


Tools for electrode handling

R
O
S
A
N
A



Connection of the electrodes



- Positioning tool with guide pins and labelling
- Connection box for distal and proximal electrodes
- Connector for 54 electrodes

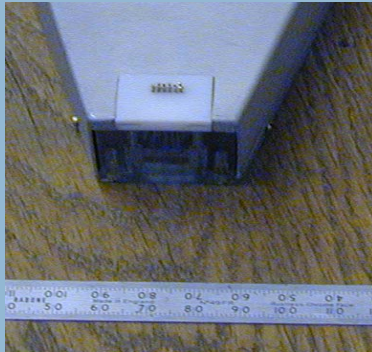
- Current or voltage controlled
- Bipolar rectangular pulse shape
- Amplitude 0.05 mA - 5 mA
0.1 V - 10 V
- Frequency 1 Hz - 60 Hz
- Pulse width 10 μ s - 500 μ s



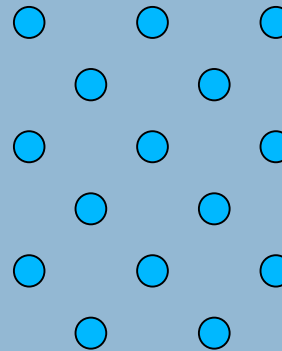
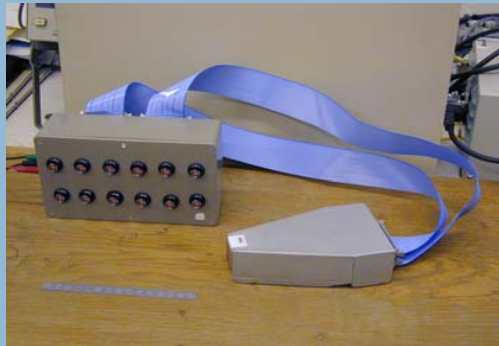
Hand held stimulator



Stimulator



- 12 pins (6 x 2 grid) spacing 1.2 mm
- 0.6 mm diameter individually retractable pins (up to 3 mm)
- Piezoelectric bimorph actuators

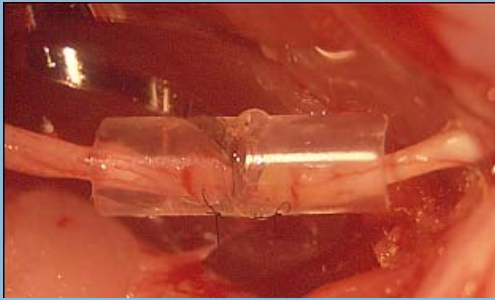


4 mm

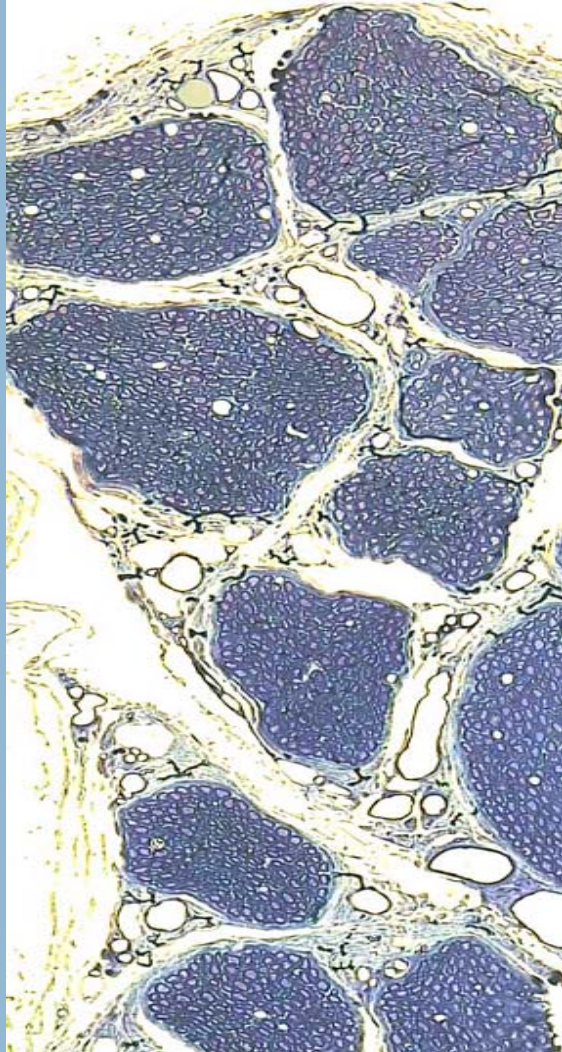
- Array of 15 pins with 1 mm row spacing
- Range of movement increased to 5 mm
- Variable drive voltage will allow variable movement



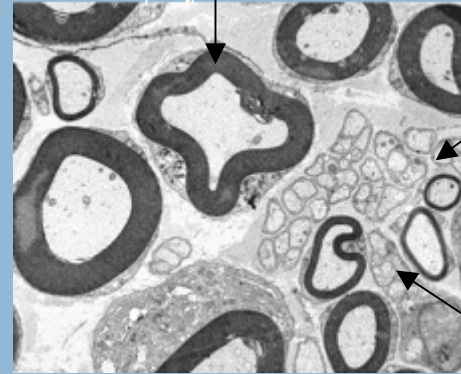
R
O
S
A
N
A



Cat median nerve (Toluidine blue)



Strongly myelinated fibers

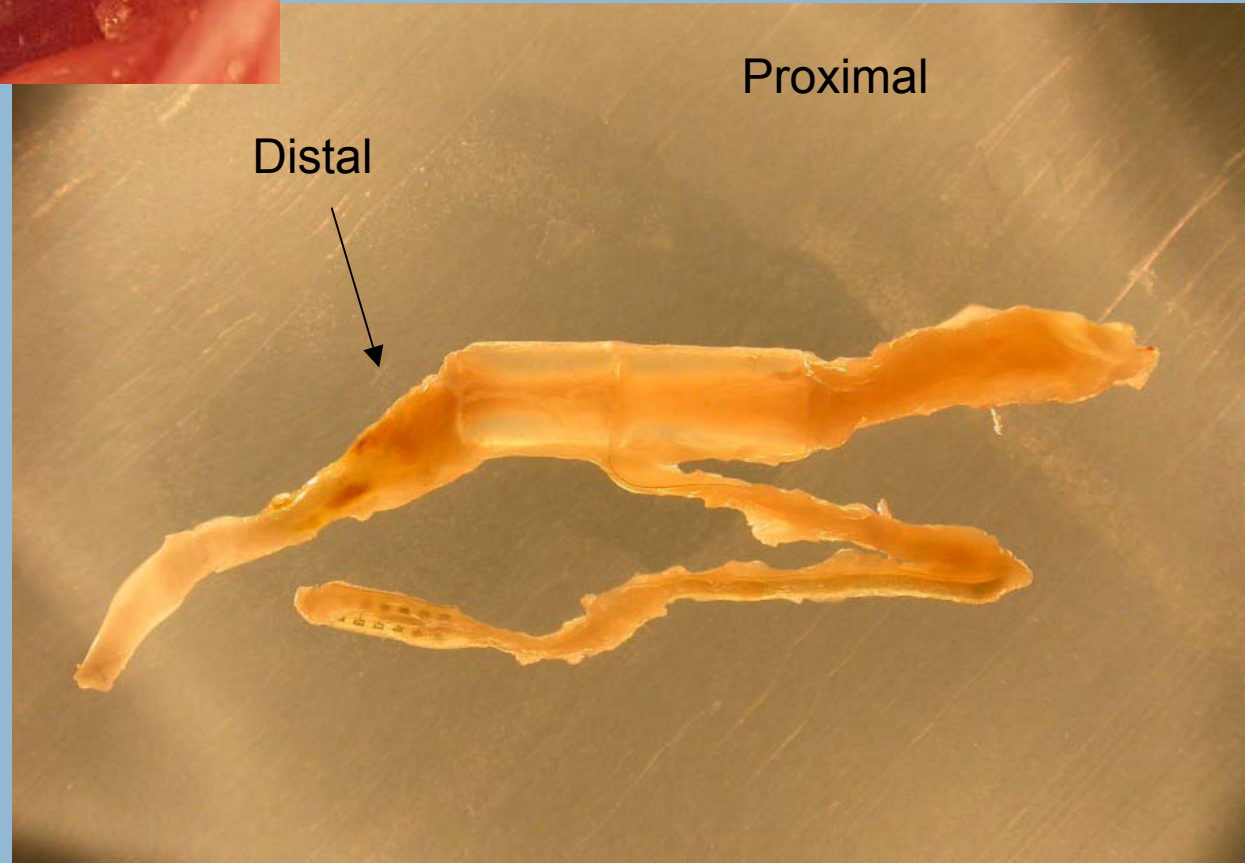
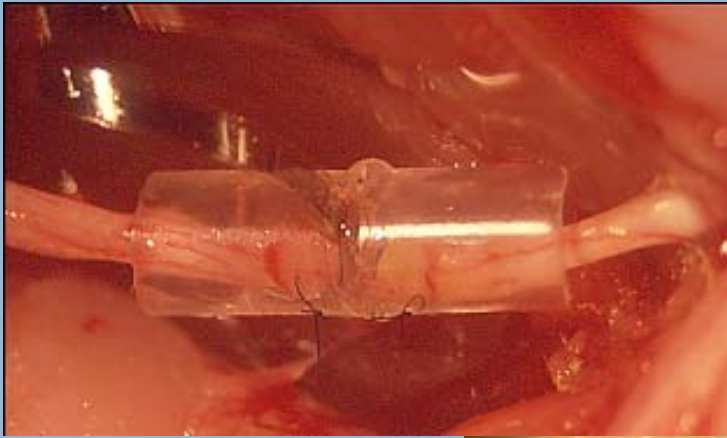


Unmyelinated fibers

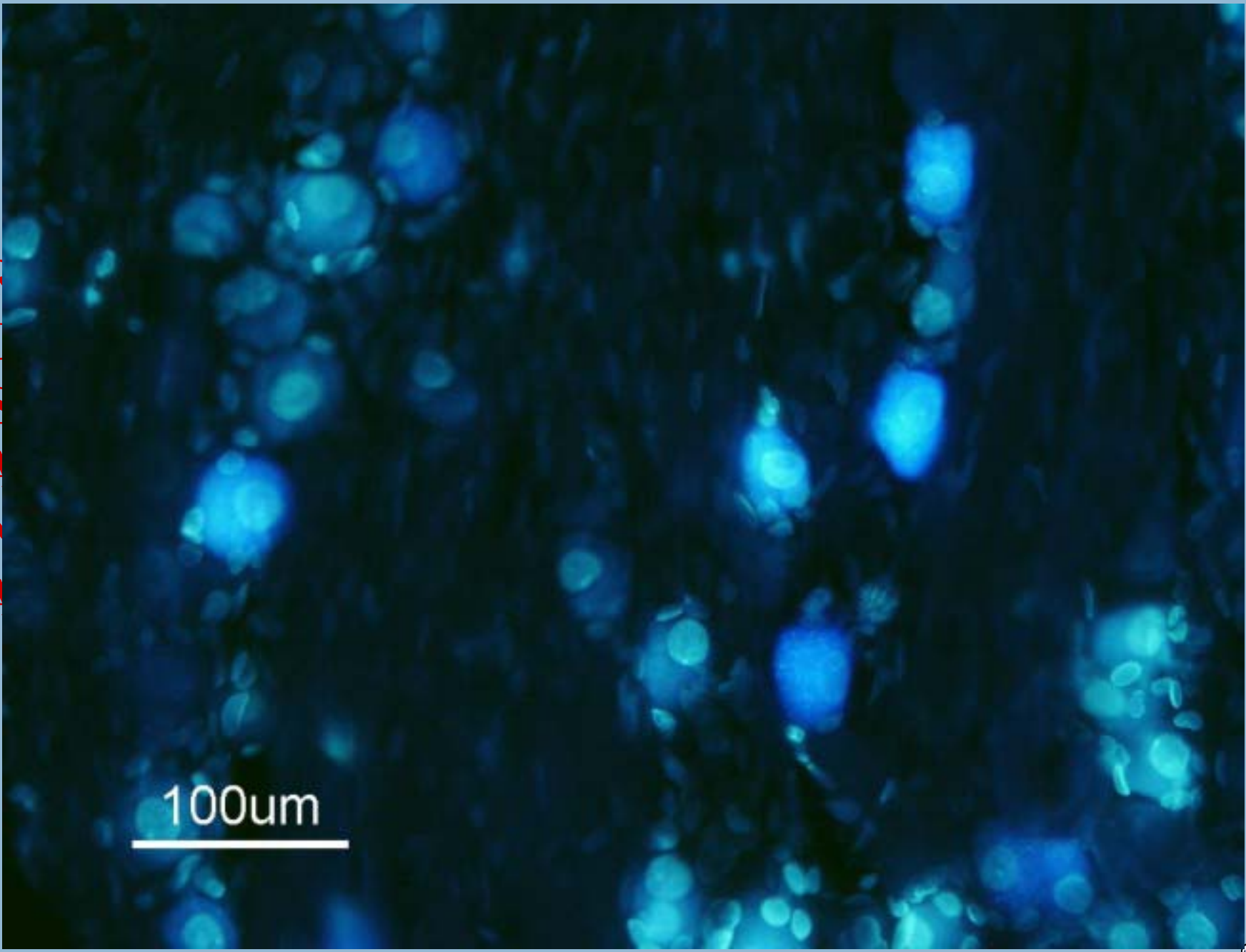
Thiny myelinated fibers



R
O
S
A
N
A

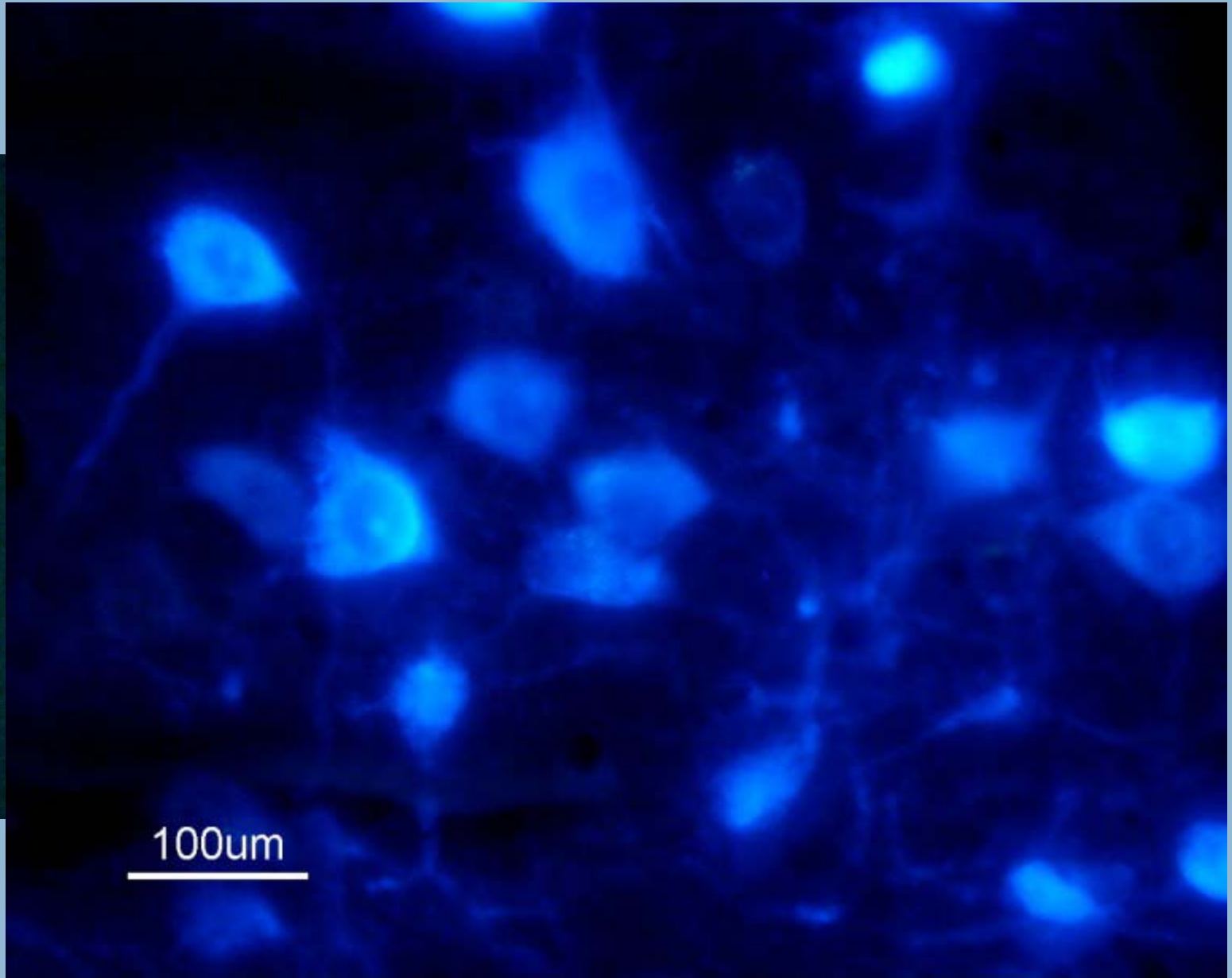
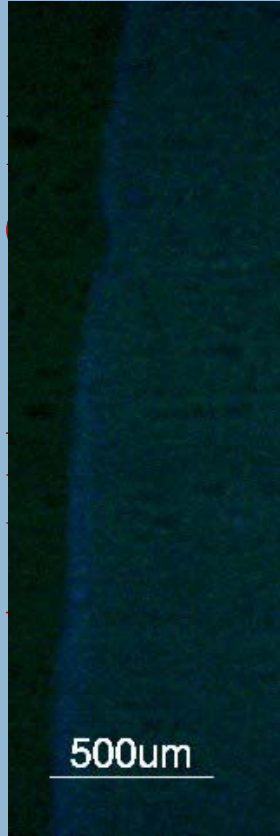


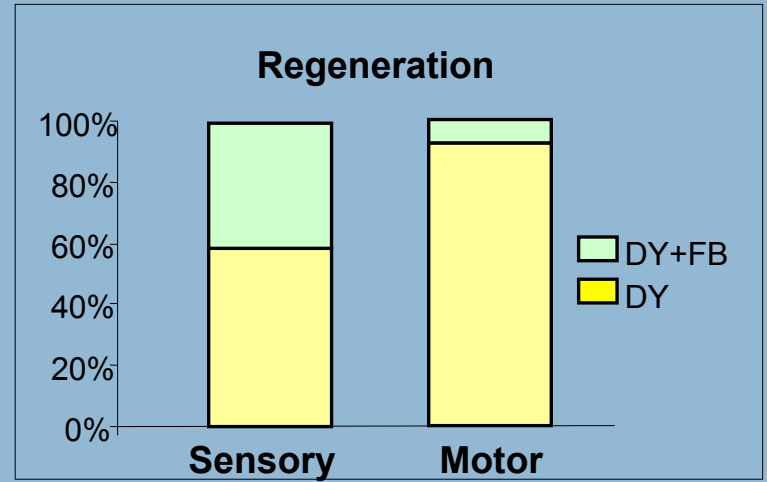
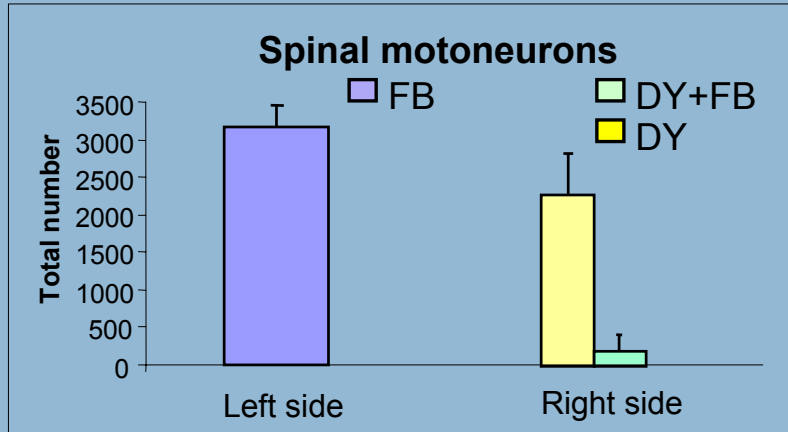
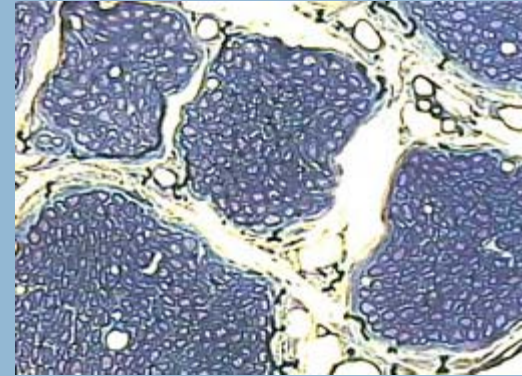
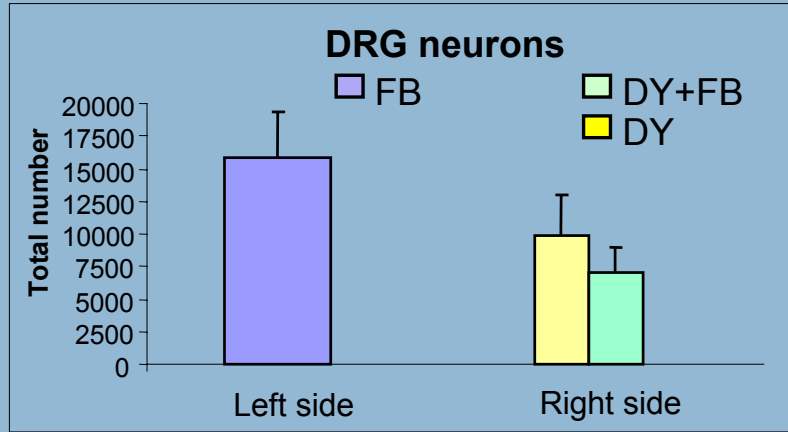
EC
CS
AN
A



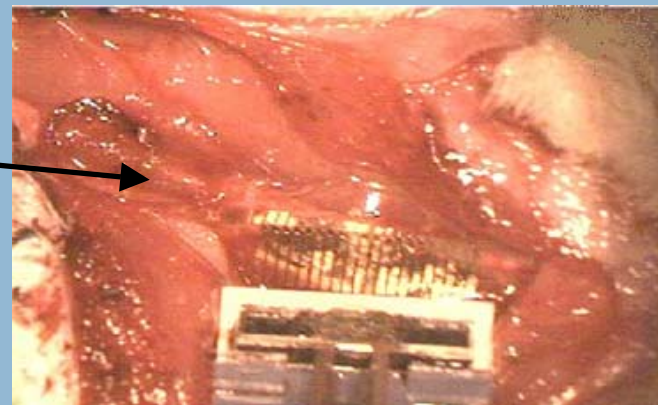
100um



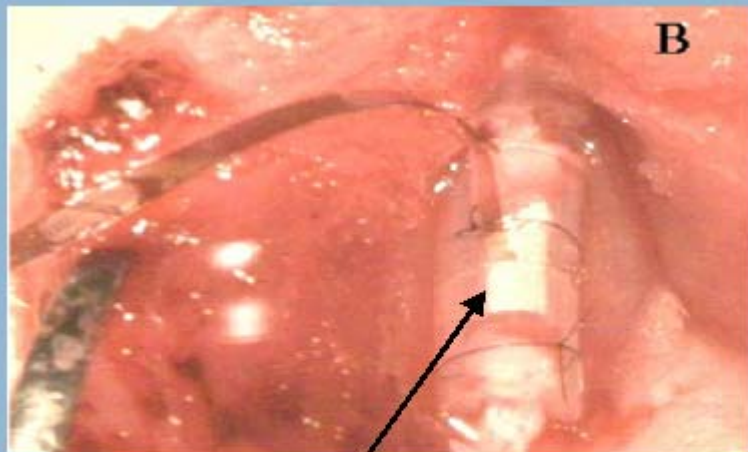




**Active Microelectrode
during an experiment**

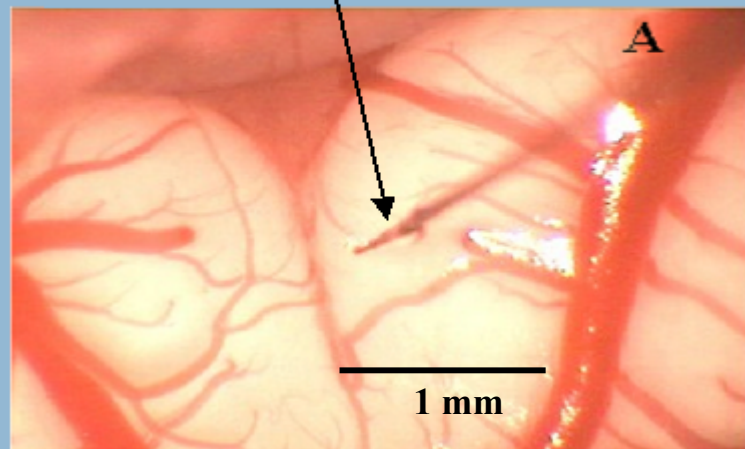


**R
O
S
A
N
A**

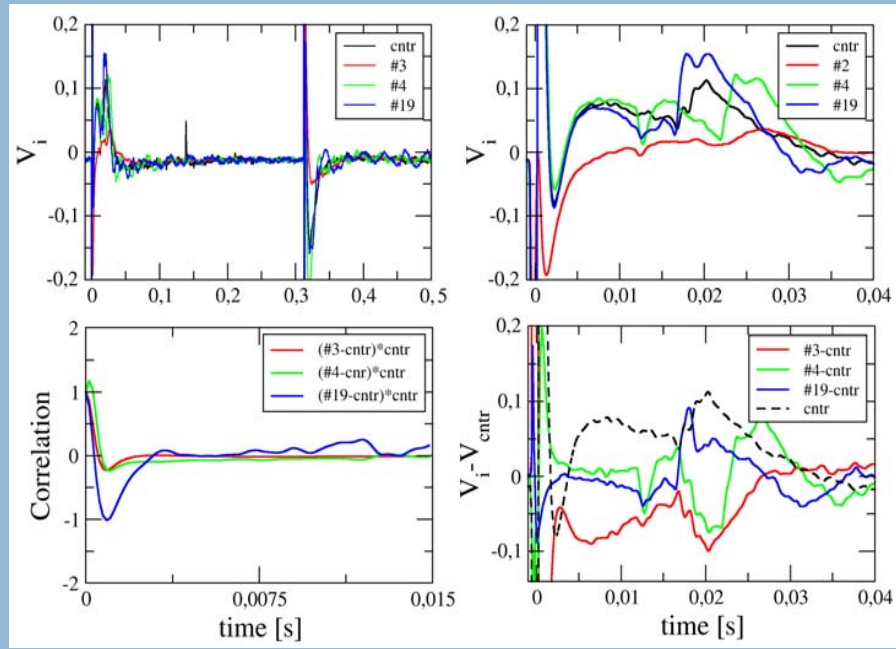


peripheral nerve recordings

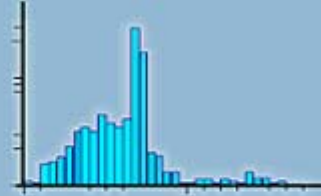
DCN recordings



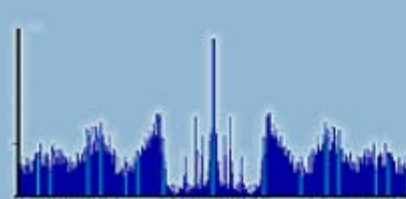
R
O
S
A
N
A



20 ms

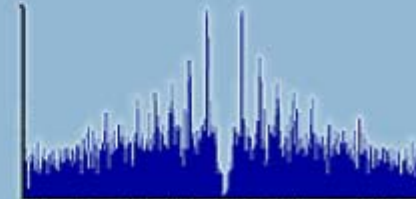


100 ms



0

500 ms



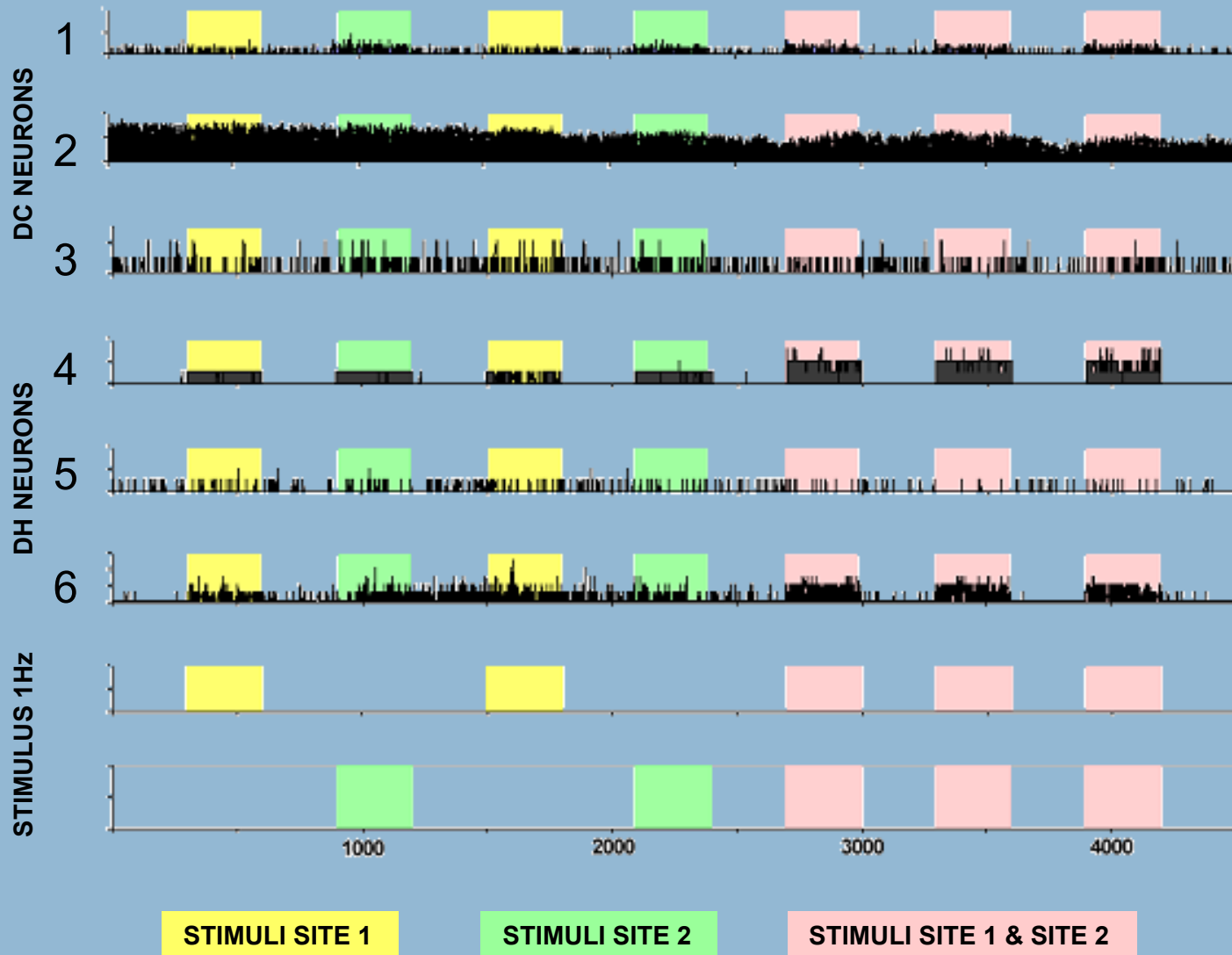
0

500 ms



RATE HISTOGRAMS

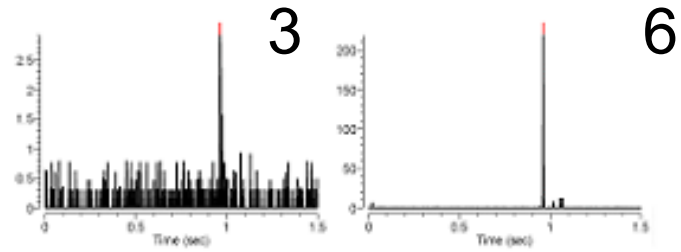
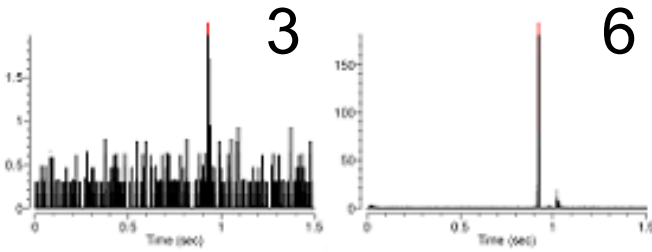
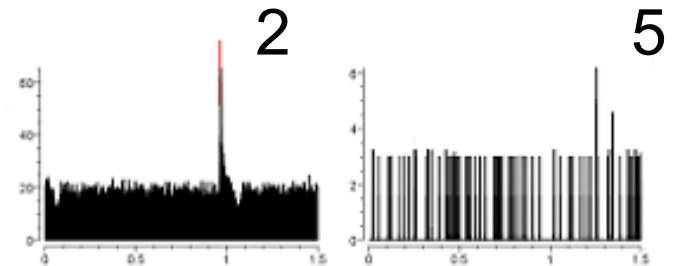
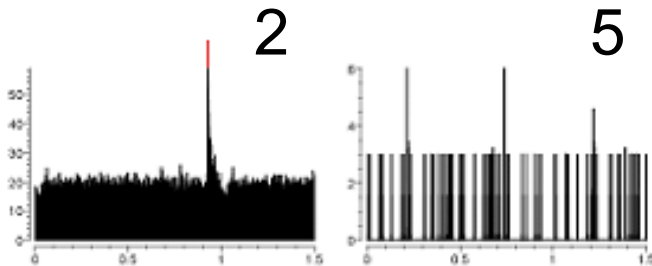
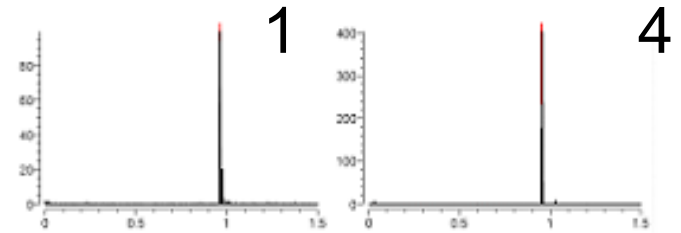
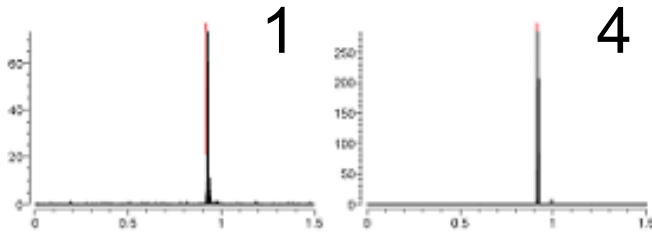
R
O
S
A
N
A



PERIEVENT HISTOGRAMS

R
O
S
A
N
A

Frequency (imp/sec)



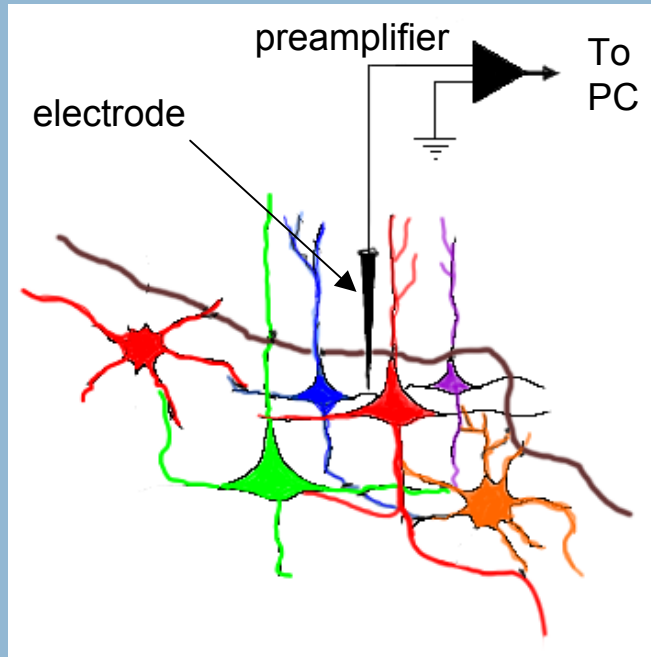
STIMULI SITE 1

STIMULI SITE 2

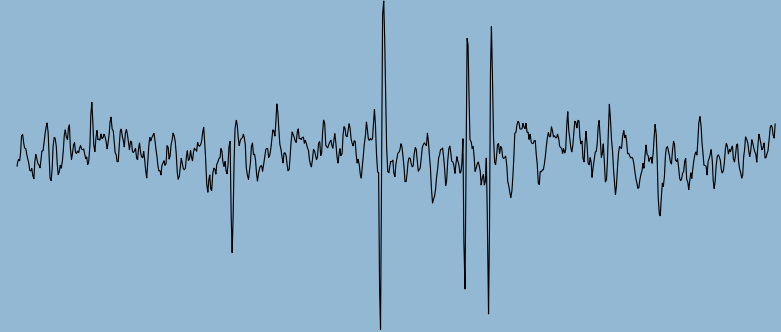


Experimental procedure

R
O
S
A
N
A



Raw experimental data



- **Simultaneous monitoring of several nearby neurons**
- **Wide band noise**
- **Detection & Separation problems**

Professional operators sorting spikes manually with tetrode electrodes do typically **from 10 to 30% errors!!!**

Performance using a single electrode is **even lower (up to 50%)!!!!**



Objective: Extract time stamps of spiking events and assign them to different neurons (the more the better)

Ideal Solution

- Takes a raw data file and produces time stamps of spiking events of prominent cells (as much as possible)
- Fast
- Efficient in memory use
- Automatic
- Accounts for spike shape variation
- Resolve spike superimposition
- Reliable!!!

R
O
S
A
N
A

- Signal denoising
- Spike extraction independent of operator mistakes
- Spike shape recovering
- Automatic spike sorting with subset resorting

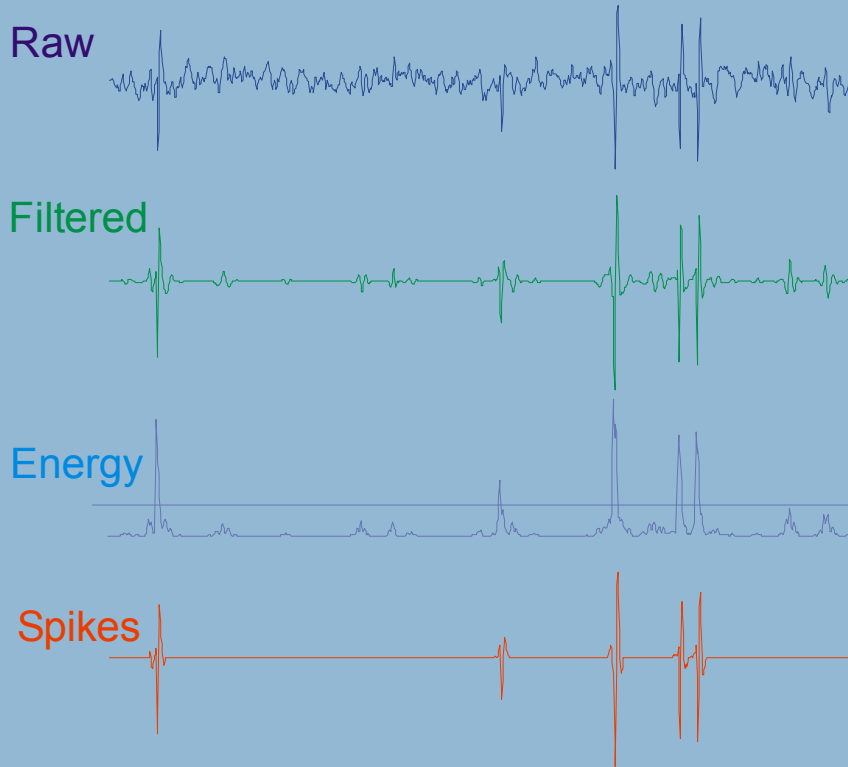
Developed packages:

- FilterSpikes (ver. 0.23*)
- DetectSpikes (ver. 0.35*)
- SortSpikes (ver. 0.52*)



FilterSpikes & DetectSpikes packages results:

R
O
S
A
N
A



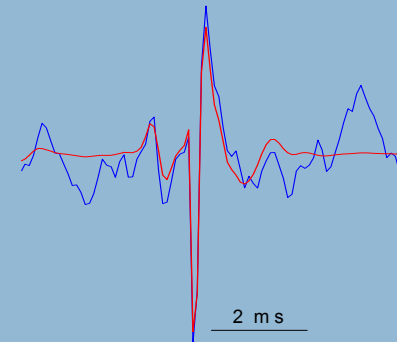
FilterSpikes

- High Pass Filtering

$F_{\text{stop}} = 400\text{Hz}$ and $F_{\text{pass}} = 600\text{Hz}$

- 2. Wavelet denoising

Undecimated DWT with hard thresholding.



DetectSpikes

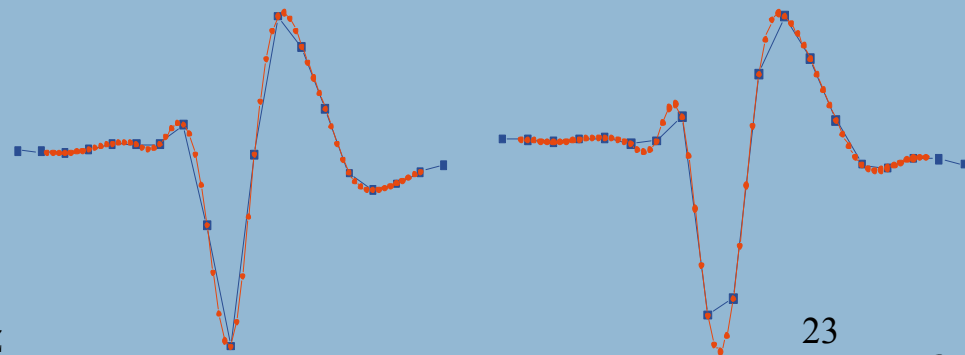
- 1. Spike extraction methods

A). Windowed Power

B). Nonlinear Energy Operator

- 2. Thresholding: $T = M + K \cdot \text{STD}$

- 3. Spike reconstruction at 40 kHz

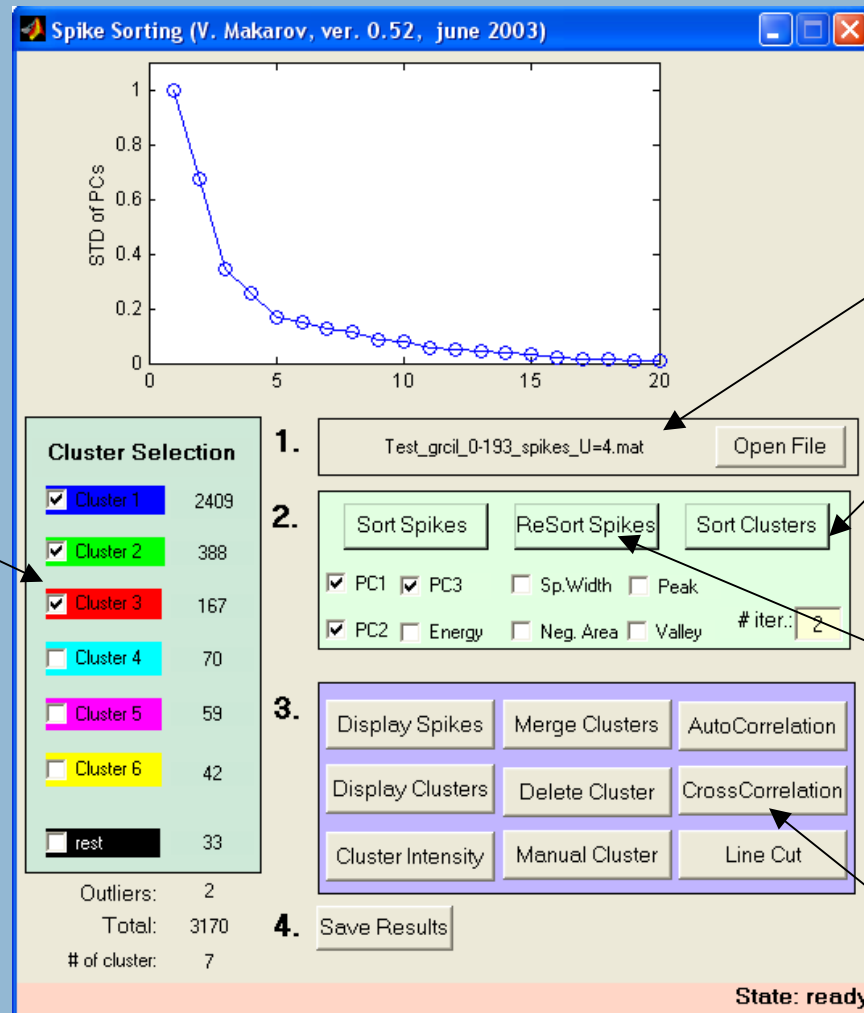


SortSpikes package

Spike waveform with 64 points at 40 kHz rate
Automatic classification with KlustaKwik
May often over split or merge clusters.

R
O
S
A
N
A

Cluster selection, color representation, and number of spikes



File with spikes and time stamps (after DetectSpikes)

Sorting buttons and features selection

Resort button, allows To perform secondary sorting on a subset of spikes

Clusters and spikes Manipulation buttons

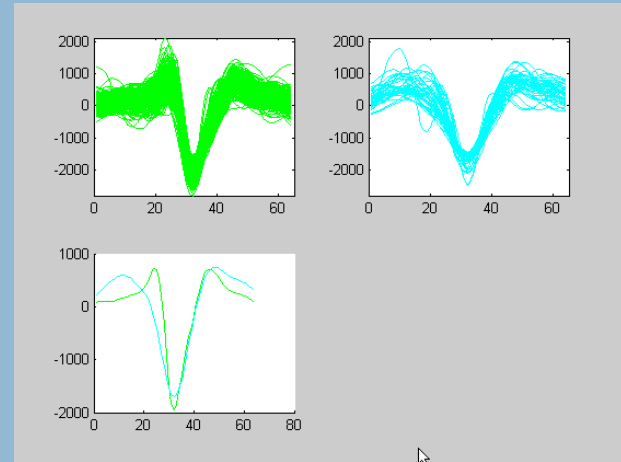
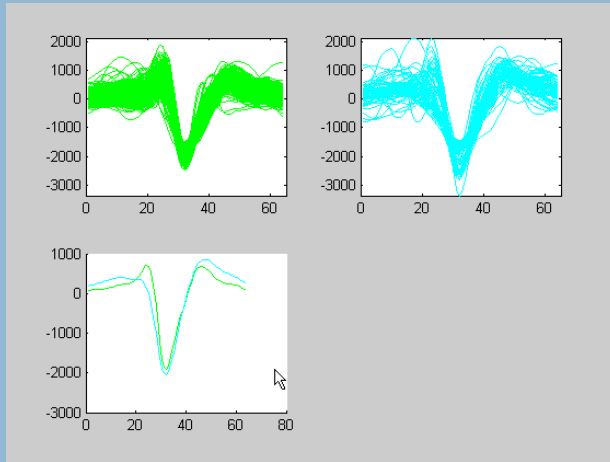


Sorting on a subset **improves** performance

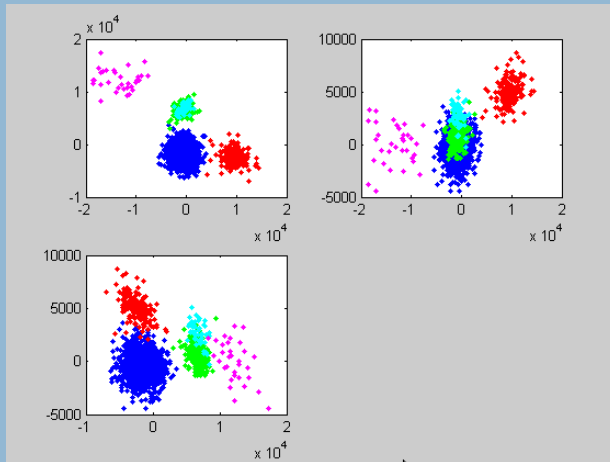
Badly separated spikes after 1st run

Improved cutting. 2nd run on **the subset**

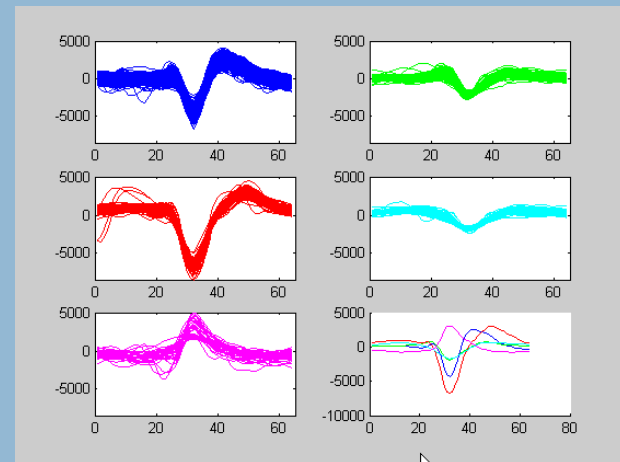
R
O
S
A
N
A



Example of final classification



Feature space



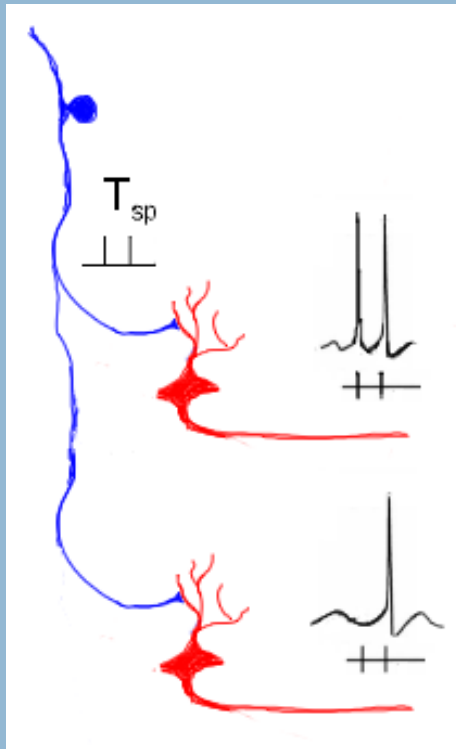
Spike wave forms



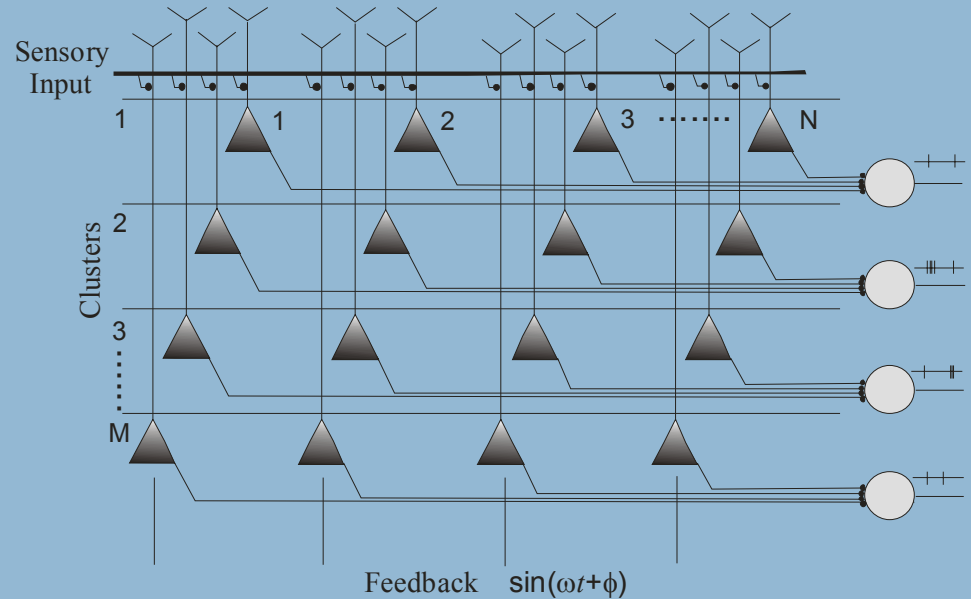
Mathematical modelling of neural networks

1. Role of subthreshold oscillations in the stimuli processing
2. Delay induced oscillatory phenomena (stability loss)





Network Organization



Minimal number of neurons in a cluster $N = \frac{1}{f_m \tau_m}$

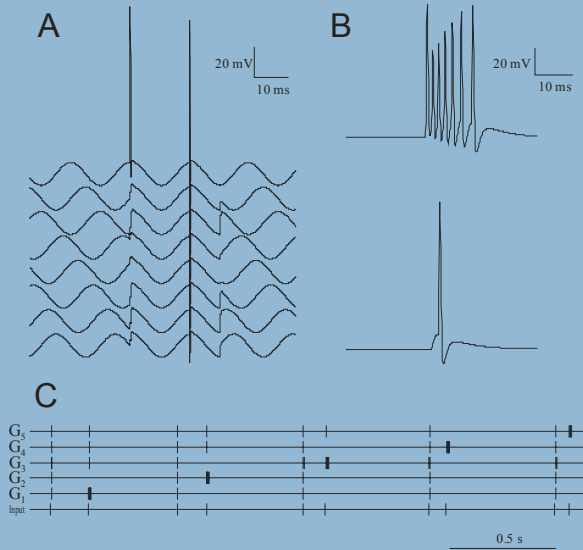
Optimal cluster frequency distribution: $f_m = \frac{(N+1)^m}{N(N-1)^{m-1}} F_{\min}$ (not linear!)

Condition for number of clusters: $\left(\frac{N+1}{N-1}\right)^M = \frac{F_{\max}}{F_{\min}}$

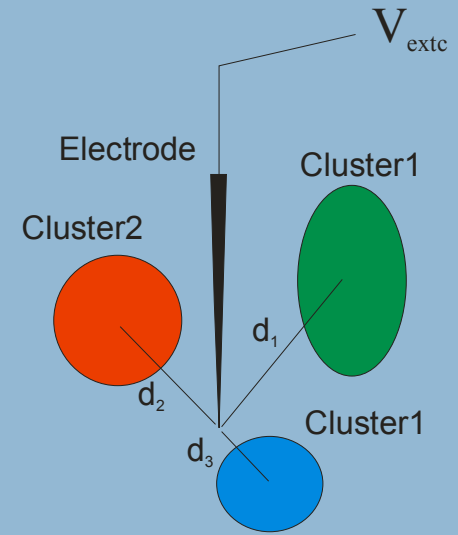
Example: For $F_{\min} = 5\text{Hz}$, $F_{\max} = 15\text{Hz}$, $N = 8$ and $M = 7$



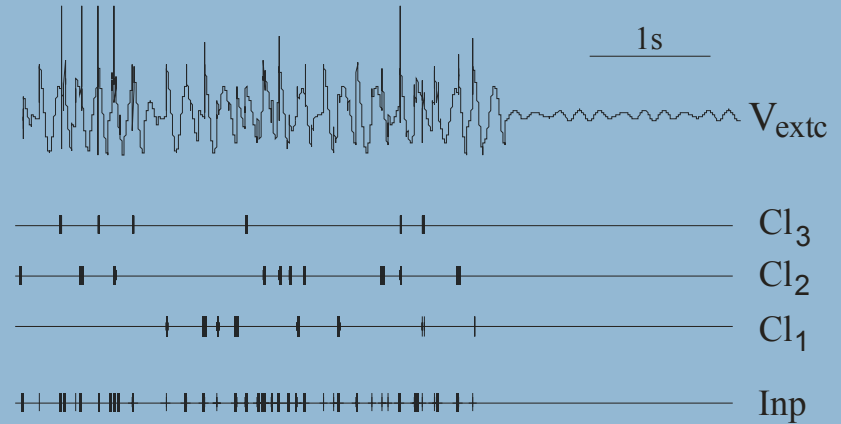
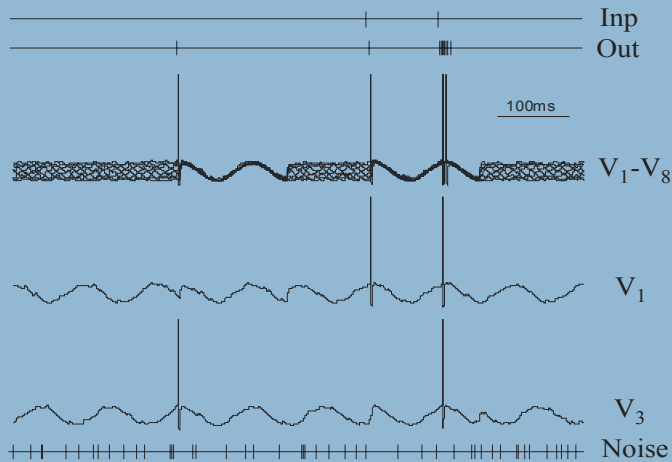
Network Response to stimulations with doublets



Synchronization of Extra-cellular activity



Robustness of interspike interval measurement

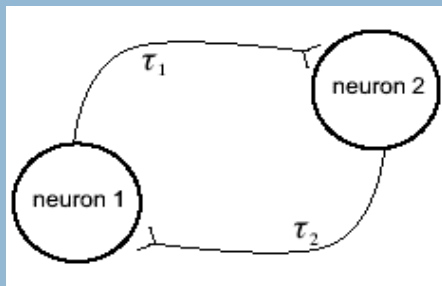


Oscillatory Phenomena and Stability of Periodic Solutions in a Simple Neural Network with Delay*

Delays occur:

1. In the signal transmission
2. In synaptic transmission

How does delay change the stability of neural network states?



Bifurcation parameters:

1. Global delay: $\tau = \tau_1 + \tau_2$

2. Composite coupling: $\nu = -abF'(0)G'(0)$

$$\dot{u}_1(t) = -\mu_1 u_1(t) + aF(u_2(t - \tau_2))$$

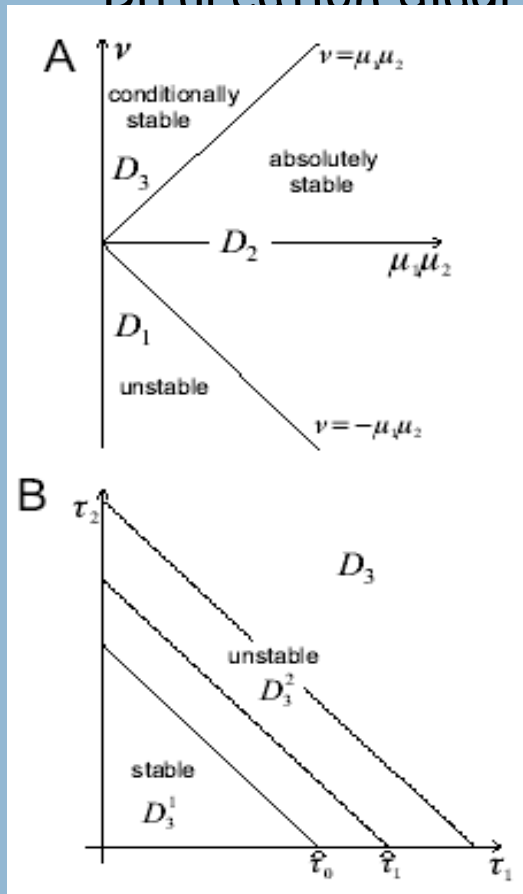
$$\dot{u}_2(t) = -\mu_2 u_2(t) + bG(u_1(t - \tau_1))$$

Positive sign - excitatory-inhibitory coupling
 Negative sign - couplings of the same type

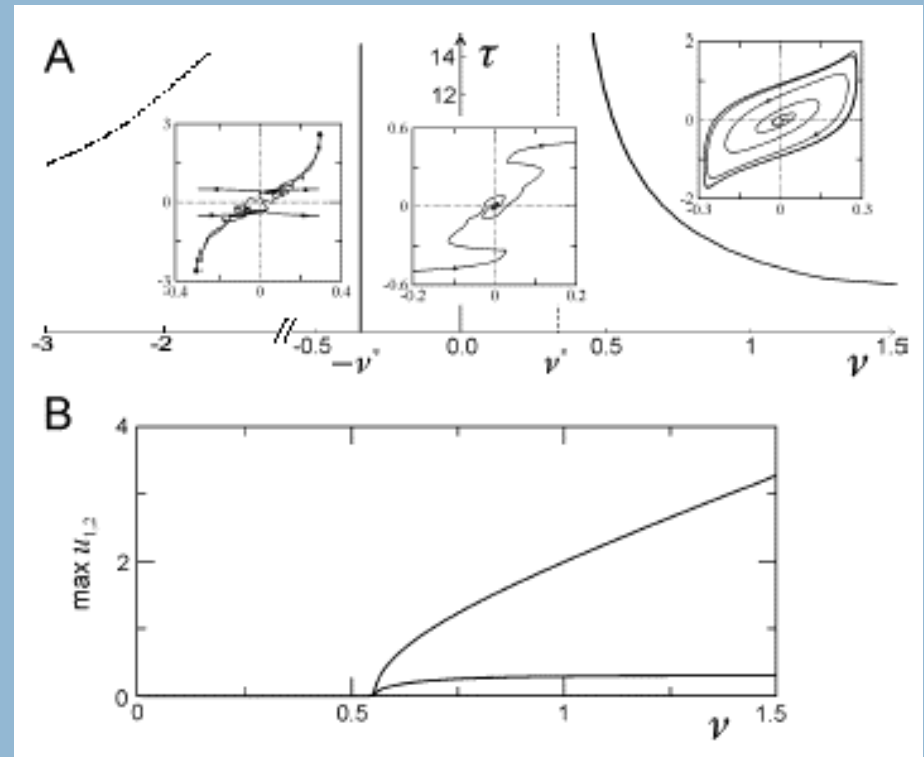
*J. Wei, M.G. Velarde, V.A. Makarov, Nonlinear Phenom. Complex Syst., 5:4 (2002) 407 - 417



Bifurcation diagram



Global existence of periodic solutions



- Delays can change the stability of neural network states
- Delay can cause delay-controlled periodic behavior
- Only composite coupling and global delay affect the system
- Periodic solution appears when the delay is large enough
- Most radical changes can occur for the excitatory-inhibitory configuration



ROSANA

R
O
S
A
N
A



Universidad Complutense de Madrid

Universidad Autonoma de Madrid

Fraunhofer Institute for Biomedical Engineering

Institute for Molecular and Cell Biology of Porto

Univ. of Exeter

